

ACEDRG

Restraint generator for monomers/ligands

Garib N Murshudov MRC-LMB, Cambridge

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- AceDRG: two functions
- Validation of entries in the DB and derived data
- Generation of new ligand description
- Covalent link description
- Identification and dealing with metals
- Conclusions

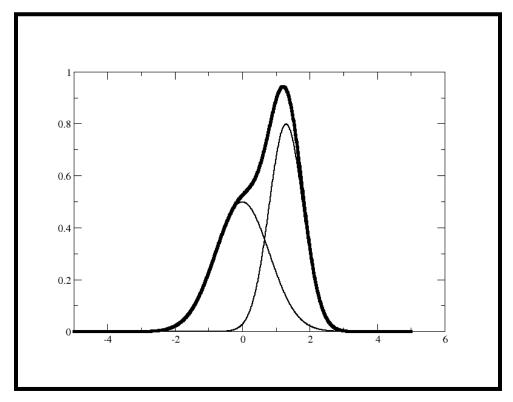
Introduction

- Information about internal structure of molecules is used to complement experimental data
- Dictionary is used for model building and fitting into the data
- Dictionary is used to position missing atoms (e.g. hydrogens)
- Properties of atoms and functional groups are used for nonbonding interactions
- Dictionary of monomers can also be used for MM calculations

Why Restraints?

Example: two-atom ideal case

Distance between atoms is 1.3Å. B-factors are 20 and 50



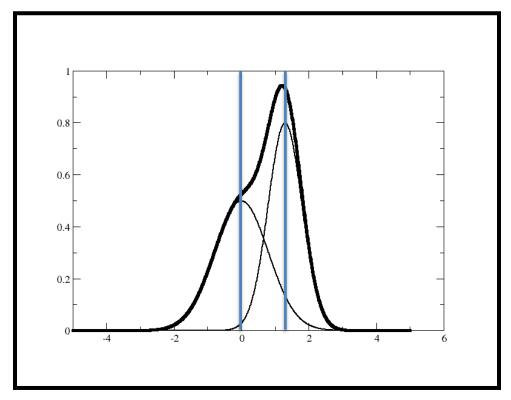
Thin lines – single atoms

Bold line - sum of the two atoms

Why Restraints?

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Distance between atoms is 1.3Å. B-factors are 20 and 50



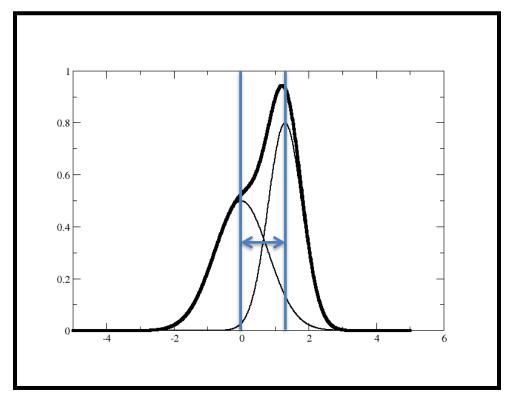
Thin lines – single atoms

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Why Restraints?

Example: two-atom ideal case

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Thin lines – single atoms

Bold line - sum of the two atoms

Restraints

Standard restraints (used by default) include:

- Bond lengths
- Angles
- Chirals
- Planes
- Some torsion angles
- B-values
- VDW repulsions

These help to ensure that the model is chemically sensible

Note – we generally deal with restraints, not constraints

Introduction

Two ways of generating chemistry of molecules:

1) Tabulated atom types and mapping of these atom types to current molecule

Problem: The number of atom types can be very large (potentially infinite)

1) Tabulated "ideal" values for building blocks of macromolecules

- 1) Monomers (e.g. AA. NA, sugars) and ligands
- 2) Links between monomers (peptide links, sugar links, NA links)

Problem: Not all building blocks have high resolution experimental structure QM may take long time if environment need to be accounted for

List of existing dictionary generators

GRADEGlobal PhasingProDRGDundee groupLibcheckCCP4Phenix.elbowPhenix groupPyrogenCOOT (Paul Emsley)AceDRGCCP4

Grade, Phenix.elbow and pyrogen use MOGUL from CSD. AceDRG at the moment uses COD as a source

Source of atom types and "ideal" values

There are two options:

- 1. High Level QM calculations
 - 1. Expensive
 - 2. It is hard to account for effects of environment
- 2. Small molecular crystallographic database
 - 1. Crystallography Open Database COD
 - 2. Cambridge Structure Database CSD

Note 1: CSD already offers very good resource – MOGUL. It is used by the programs GRADE, phenix and coot's pyrogen.

Note 2: All small molecular databases have their own problems: data must be validated before use

AceDRG: two functions

We need to solve two problems

- 1. Derive atom types from small molecular database
- 2. Apply derived parameters to generate new ligand description

Both of these problems are dealt with AceDRG. It generates parameter tables and applies them to new ligands.

Atom types are dynamic: if database is extended or new database is used then AceDRG can be applied to generate new parameter tables

AceDRG applied to COD

ACEDRG currently uses small molecule structures from Crystallography Open Database(COD) as a source for:

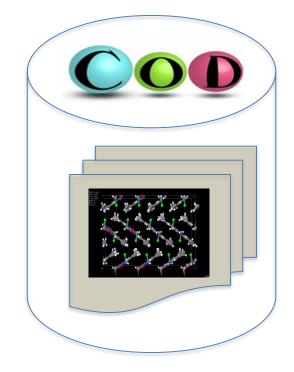
□ Atom types that characterize atom's local environments.

Derivation the values of bond lengths and angles

Data Source

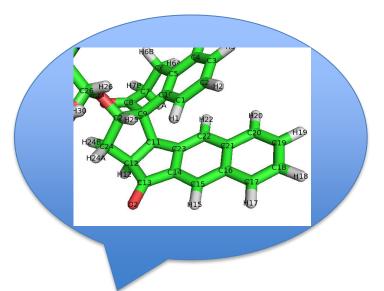
Brief summary of COD

- COD contains more than 300,000 crystal structures of organic, inorganic and metalorganic compounds. (for comparison, number of ligands in PDB : ~23000)
- All structures are published in peerreviewed journals.
- It is freely accessible and updated daily.



Rules for atom types

- 1. Element name
- 2. The number of bonds
- 3. Belongness to rings (up to 6 member)
- 4. Aromaticity of these rings
- 5. Information about immediate neighbours
- 6. If atoms are in aromatic rings then some information about their third neighbours



Atom C23: C[5,6a](C[5,5]C[5,5]C[5,6]H)(C[5,6a]C[6a]C[5])(C[6a]C[6a,6a]H) Atom C18: C[6a](C[6a]C[6a,6a]H)(C[6a]C[6a]H)(H)

Rules for hybradisation

sp³:

- 1. C, N, B with four bonds
- 2. S with three or four bonds
- 3. O with two bonds and none of the bonded atoms is sp²

sp²:

- 1. C with three bonds
- 2. N or B with three bonds and one of the attached atoms is sp^2
- 3. O with two bonds and one of the attached atoms is sp²
- 4. O with one bond

AceDRG applied to COD

Curating and Validating data from COD

- Total structures in COD:
- Rejected structures :

307987

175923

Resolution is low (>0.88A) or Rfactors are high (> 0.10) and no symmetry operator, occupancies ≠ 1 : 72067

Rejected by basic chemical sanity check:

- the number of bonds to an atom should be consistent with its valence
- Difference between same type of bonds within molecule should not be very different: 103856

AceDRG applied to COD

Curating and Validating data from COD

Extended organic Set Includes: H, C,N, O, B,P, S, F Cl, I, Br, Se

Structures with metals as first or second neighbour: **7286**

Number of COD structures used by AceDRG:12477840% of COD

Validation & Quality Control

COD:

Crystallographic & chemical validation

ACEDRG:

Crystallographic & chemical validation (use CIF files from COD)

Separate Validation Tool:

Systematic validation Statistical analysis

Coot:

Final ligand validation

- checks consistency against reference
- can use multiple sources acedrg (COD), pyrogen (Mogul/CSD)
- resource comparison

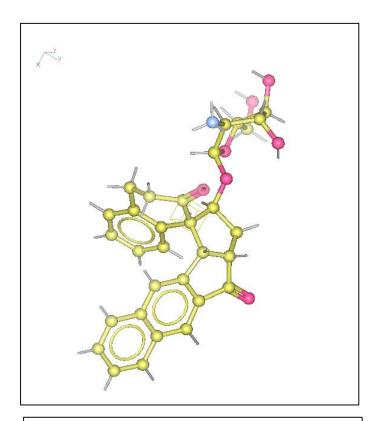
AceDRG as a dictionary generator

Monomer/Ligand/restraint generator

- Input files of SMILES, SDF/MDL, mmCIF formats
- Output Dictionary files of mmCIF format: contain atom types, bond lengths and angles, torsion angles, planes and chiral centers
- Output coordinate files of PDB format, which represent one of the conformations.

Using Acedrg

RDKit is used to accept input files of different formats: SMILES, MDL/SDF mol, SYBL/MOL2. MMCIF files are converted to mol files first.



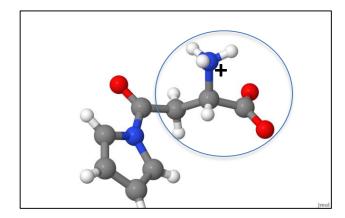
N[C@@H]1[C@@H](O)[C@H](O)[C@@H(CO)O[C@H]1O[C@H]2C[C@H]3[C@H](c4 cc5cccc5cc4C3=O)[C@@]26C(=O)CCc7cc ccc67

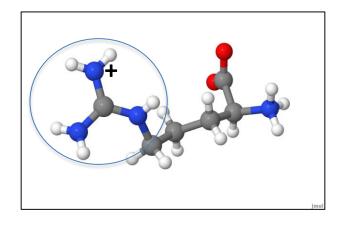
Functional groups in AceDRG

There are 21 functional groups with their assosiated pKa values. Functional groups are used to make decision about protonation state of a ligand. Current assumption is that pH is 7.0. But it can be changed by a user or software running acedrg.

The list of functional groups can be extended.

Functional Groups in Acedrg: example



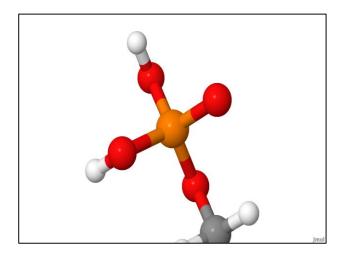


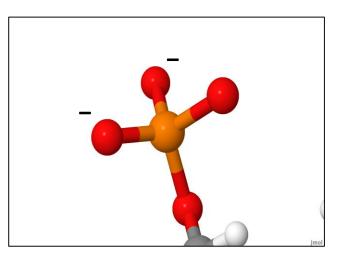
Substructure in ASM pKa : 2.0 PH : 7.0 Substructure in ARG pKa: 12.5 PH : 7.0

The default pH is 7.0

Functional Groups in Acedrg: example

Phosphate group PO4 in e.g. PYO





In PDB: in vacuum?

In Acedrg, PH=7.0

Use of Acedrg

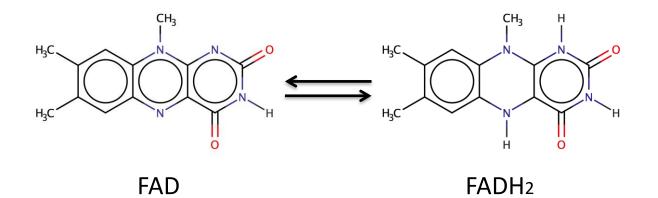
Ligand/restraint generator

- Input files of SMILES, SDF/MDL, mmCIF, SYBYL/Mol2 formats
- Output monomer/ligand files of mmCIF format, which contain atom types, bond lengths and angles, torsion angles, planes and chirality centers, and are used as restraint files for refinements.
- Output coordinate files of PDB format, which represent one of lower energy conformations for the ligands under consideration.

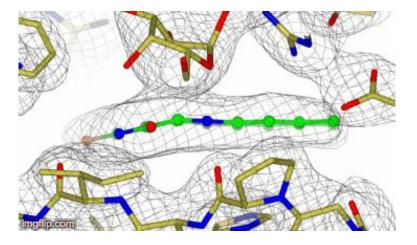
An example: Effect of aromaticity definition

AceDRG

An example of FADH₂ with two different aromaticity definition



PDB 3HDY FADH₂ before and after refinement with AceDRG-generated restraints



Using Acedrg in ccp4i2

In CCP4i	2 interface
	+ CCP4-7.0.077 Project Viewer: hmm
Task menu Export project Run Run on server Clone job Help	Bibliography Export MTZ Show log file
Job ist Project directory Filter: Only show jobs containing text typed here	
Job/File Evaluation 1 Make Ligand - Acedrg 	
• I make Liyanu - Aceury	Job title Make Ligand - Acedrg
	Start point Start with molecular structure from a sketch Will launch Lidia to sketch molecule. Click Apply and Close in Lidia when sketch is ready. Optionally can provide a starting monomer for the Lidia sketch: Image: Click Apply and Close in Lidia when sketch is ready. Optionally can provide a starting monomer for the Lidia sketch: Image: Click Apply and Close in Lidia when sketch is ready. Optionally can provide a starting monomer for the Lidia sketch: Image: Click Apply and Close in Lidia sketch:
	Three letter code for output monomer DRG
	Atom Naming Attempt to match atom names with nothing

Using Acedrg in ccp4i2

In CCP4i2 interface

				X	Lidia: Co	oot's Ligar	nd Builder						
<u>F</u> ile Di	splay <u>H</u>	elp											-7.0.077 Project Viewer: hmm
e Clear	🥱 Undo	— Single	= Double	≡ Triple	↓ Stereo	💋 Charge	Cut	Delete	Hydrogens) SMILES	J. Tidy Up	•	phy Export MTZ Show log file
 3-C	□ 4-C	D 5-C	0-C	O 6-Arom	0 7-C	0 8-C	Env. Res		Key				Make Ligand - Acedrg The job is Pending The job is
C N S P H F Cl Br I X		[рн J									Use data from job No o as input below point with molecular structure from aunch Lidia to sketch molecule. Click Apply and Close in Lidia when sketch is ready. anally can provide a starting monomer for the Lidia sketch: Mol fileis not used t monomer letter code for output monomer DRG Naming pt to match atom names with nothing
										C c c	<u>•</u>	_	
SMILES:	0CC1CC	CC(CI)C1									le 1.00 Show Ale		

Covalent Links in AceDRG

Description how to use AceDRG can be found here:

https://www2.mrc-lmb.cam.ac.uk/groups/murshudov/content/acedrg/acedrg.html

Example: Create a file containing something like this:

LINK: RES-NAME-1 2OP FILE-1 2OP_acedrg.cif ATOM-NAME-1 C RES-NAME-2 VAL ATOM-NAME-2 N

Write to a file. For example – myInsts.txt

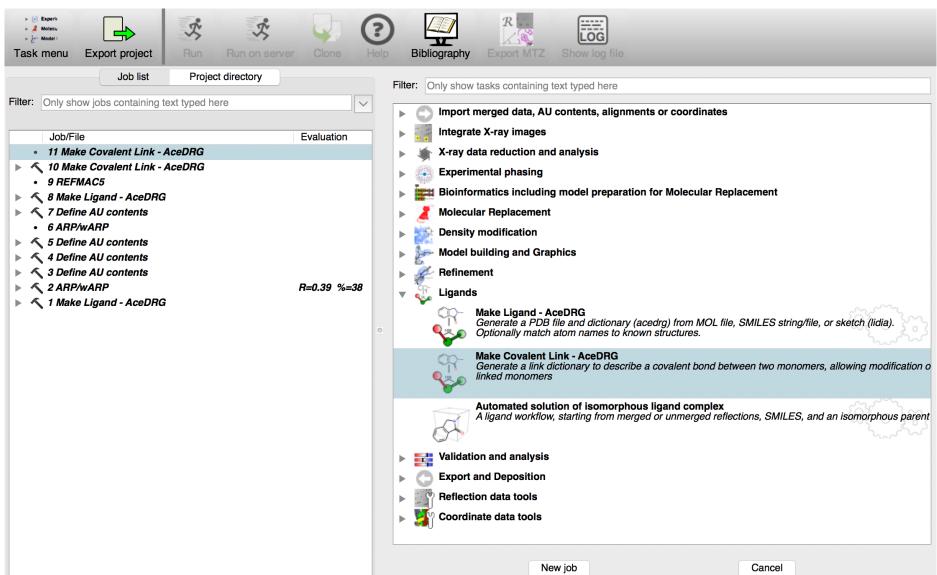
acedrg -L myInsts.txt

It will create files corresponding covalent links.

This option is not yet available on the interface. It will be available soon.

Covalent links using ccp4i2

From task menu select "Make Covalent Link – AceDRG"



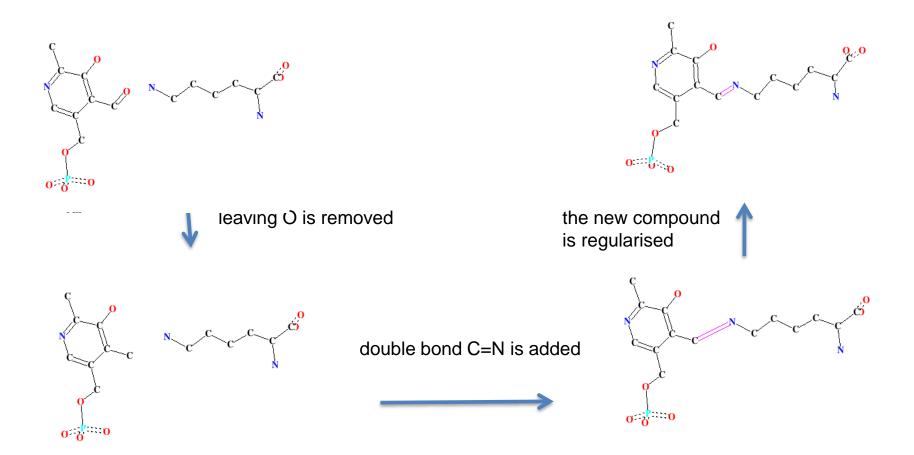
Covalent links using ccp4i2

Job list Project directory	·	Job 11: Make Covalent Link	- AceDRG	The job is Pending	
ilter: Only show jobs containing text typed here	~		Input Result	ts Comments	
	·		Input data	Advanced	
Job/File	Evaluation				
• 11 Make Covalent Link - AceDRG		Job title Make Covalent Link - Ac	eDRG		
 10 Make Covalent Link - AceDRG 9 REFMAC5 		First monomer to be linked			
▶ ≪ 8 Make Ligand - AceDRG		Get ligand description from the	Monomer Library		
 7 Define AU contents 6 ARP/wARP 		Residue name PLP linking at	om C4A 🔅		
5 Define AU contents		Delete atom O4A			
► < 4 Define AU contents					
 3 Define AU contents 3 2 ARP/wARP 	R=0.39 %=38	Change order of bond			
 ZARP/WARP A 1 Make Ligand - AceDRG 	R=0.39 %=38	Second monomer to be linked			
		Get ligand description from the	Monomer Library		
		Residue name LYS linking at	om NZ 🗘		
		Delete atom			
		Change order of bond			
		Order of the bond between linked	atoms double 🗘		
		Apply links to model (optional)			
		Atomic modelis no	tused		
		Create links between all mate	hing atom-pairs		

Creating a new link, as seen in JLigand GUI

The two monomers are in effect reacted in silico Hydrogen atoms are dealt with automatically^{*)}

*) it is also possible to visualise H-atoms and deal with them explicitly



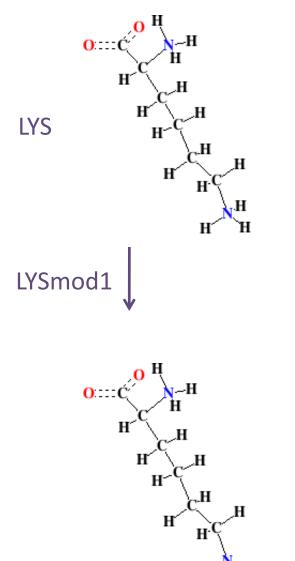
Both monomers are modified and link is created

Contents:

- (1) modification "PLPmod1"
- (2) modification "LYSmod1"
- (3) link "PLP-LYS"

	000	PLP-LYS			
	data_mod_list				
F	loop_ _chem_mod.id _chem_mod.name _chem_mod.comp_id _chem_mod.group_id PLPmod1 "PYRIDOXAL-5'-PHO LYSmod1 'LYSINE data_link_list	DSPHATE '	" LYS .	PLP .	0
∠ 3`	loop_ _chem_link.id _chem_link.comp_id_1 _chem_link.group_comp_1 _chem_link.comp_id_2 _chem_link.mod_id_2 _chem_link.group_comp_2 _chem_link.name PLP-LYS PLP PLPmod1 . PLP-LYS	LYS	LYSmod1 .		
				Close)
				_	-

The new link, "file view"



Modification "LYSmod1":
changes to LYS

data_mod_LYSmod1

.

loop_ _chem_mod_atom.mod_ _chem_mod_atom.functi _chem_mod_atom.new_a _chem_mod_atom.new_t _chem_mod_atom.new_t _chem_mod_atom.new_p LYSmod1 change NZ LYSmod1 delete HZ1 LYSmod1 delete HZ2 LYSmod1 delete HZ3	on id atom_id ype_symbol ype_energy partial_charge N 	0.000		Atoms
loop_ _chem_mod_bond.mod_ _chem_mod_bond.functi _chem_mod_bond.atom_	on _id_1			
_chem_mod_bond.atom_ _chem_mod_bond.new_t				
_chem_mod_bond.new_v _chem_mod_bond.new_v				
LYSmod1 change CE LYSmod1 delete NZ H LYSmod1 delete NZ H	NZ . 1Z3 .	1.455	0.020	Bonds
LYSmod1 delete NZ	IZ1 .			

Angles

Utilising new link description

Three remaining steps:

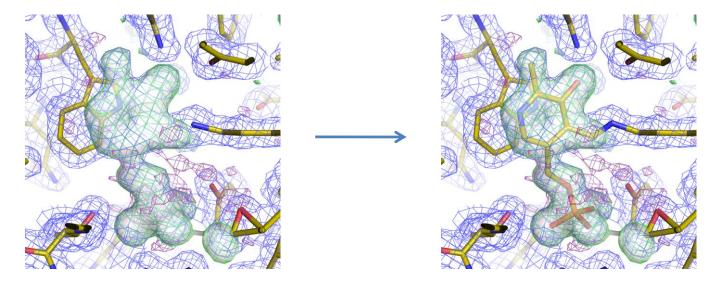
- docking monomer(s) into electron density
- defining link in the pdb-file
- refinement of the structure with linked ligand using additional library

(1) Docking into the electron density

In our example, this is completely independent step: the additional library is not used.

- non-modified monomer is taken from the standard library

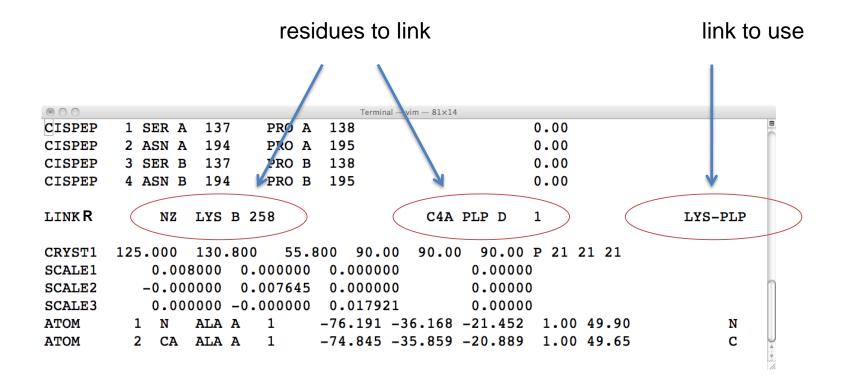
- docking is performed, e.g. using coot:



- leaving atoms (O4A of PLP in this example) are removed

(2) Defining link in the pdb-file

Ccp4i2 should be able to edit the pdb/mmCIF file and add link record



Metals

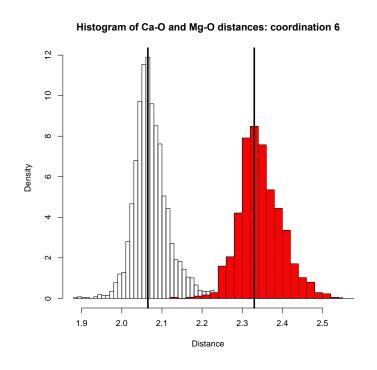
Metal identification

- 1. Distance to coordinating atoms
- 2. Coordination numbers
- 3. B values
- 4. Difference maps
- 5. Anomalous difference maps

Metals

Mg (black histogram) and Ca have definite coordination distance to O. For Ca: distance is around 2.34 For Mg: distance is around 2.07

Actual distance will depend on surrounding stoms

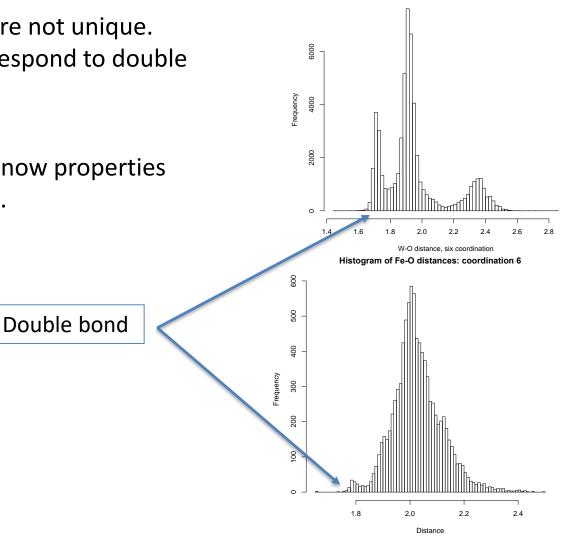


Red – Ca O distance: coordination 6 Black – Mg O distance coordination 6

Not all metals have potential distances

W and Fe and O distances are not unique. One of these distances correspond to double bond covalent.

It would help if you would know properties of the metal in your protein.



Histogram of distances between W and O

Some of the links related to metals

ZN-CYS

ZN-HISND

ZN-HISNE

FE-CYS

FE-HISND

FE-HISNE

All these and many other links are available from the monomer library. You can access them using and editor and view the file

\$CLIBD_MON/list/mon_lib_list.cif

There are few more pretabulated links related to HEC-CYS and others

Conclusions and future perspectives

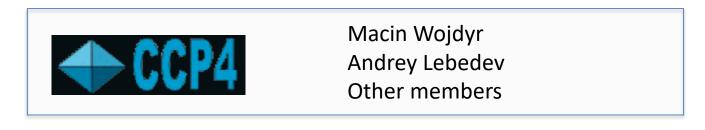
- Local graph based dynamic atom typing was designed
- Crystal structures from COD are validated
- AceDRG can be used to derive data from any small molecule database
- COD is currently used, but CSD or other databases can be used
- Derived data and AceDRG are available from CCP4
- Metals can be identified using distances, B values, difference maps, anomalous difference maps

Future

- Use AceDRG tables for ligand validation
- Deal with metal containing ligands (proteins)
- CCP4i2 interface for links and metals

Acknowledgement

MRC Laboratory of Molecular Biology	Fei Long Paul Emsley Rob Nicholls	
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Jon Agirre